**Introduction**

SS is an inflammatory autoimmune disease, with focal lymphocyte infiltration and inflammation in exocrine glands. In SS, lymphocytes selectively target moisture-producing glands, especially the salivary and lacrimal glands, with subsequent loss of their ability to produce saliva and tears, resulting in dryness of the mouth and eyes (xerostomia and xerophthalmia, respectively). Some of the studies have suggested that Toll-like receptors and B cells play a pivotal role in the pathogenesis of autoimmune diseases such as systemic lupus erythematosus, rheumatoid arthritis and SS etc. Stimulation of B cells via the TLRs pathway leads to several important changes including increase in antibody production, differentiation to plasma cells, cytokine production and up-regulation of molecules essential for antigen presentation to (autoreactive) T cells. Experimental data support the idea that co-engagement of BCR and TLR might be sufficient for B cell activation and lead to the failure of tolerance. In human naive B cells, most TLRs are expressed at very low or undetectable level, but expression of TLR 7 and 9 is rapidly induced by B cell receptor triggering. This review will focus on the possible role of B cells and TLRs signaling in the pathogenesis of SS.

**Summary:** Sjögren’s syndrome (SS) is a chronic autoimmune immunopathological disease of unknown aetiology. It is characterized by focal lymphocyte infiltration and inflammation in exocrine glands, involving especially salivary and lacrimal glands. Hypofunction of the glands leads to the decreased glandular secretion together with impaired production of saliva and tears, resulting in dryness of the mouth and eyes (xerostomia and xerophthalmia, respectively). Some of the studies have suggested that Toll-like receptors and B cells play a pivotal role in the pathogenesis of autoimmune diseases such as systemic lupus erythematosus, rheumatoid arthritis and SS etc. Stimulation of B cells via the TLRs pathway leads to several important changes including increase in antibody production, differentiation to plasma cells, cytokine production and up-regulation of molecules essential for antigen presentation to (autoreactive) T cells. Experimental data support the idea that co-engagement of BCR and TLR might be sufficient for B cell activation and lead to the failure of tolerance. In human naive B cells, most TLRs are expressed at very low or undetectable level, but expression of TLR 7 and 9 is rapidly induced by B cell receptor triggering. This review will focus on the possible role of B cells and TLRs signaling in the pathogenesis of SS.

**Key words:** Sjögren’s syndrome; Pathogenesis; B cell; TLR; Activation
are women. The exact prevalence is unknown, because many of its milder forms elude diagnosis. The prevalence is estimated to be about 0.6%, according to the criteria used for diagnosis (55). The precise etiology as well as causal therapy is at present unknown and current treatment is mainly palliative (19).

It is supposed that the etiology of SS is multifactorial. Some combination of genetic, hormonal, viral or environmental factors may be involved in the pathogenesis. Female predominance appears to be due to the ability of sex hormones to modulate immunity. Estrogens are known to be natural immune enhancers, whereas androgens are immune suppressors (37). Estrogen deprivation may play a role, as the disease often develops after the menopause. Moreover, Sjögren’s syndrome-like disease often develops in estrogen-deficient mice (33). The association of Sjögren’s syndrome with human leukocyte antigen (HLA)–B8, HLA-DR3, and with the DQA1*0501 allele suggests the idea of genetic predisposition (43). Study from Ivanyi et al. as one of the first confirmed significantly higher frequency of HL-A8 in patients with SS (16). Viral infection as a possible cause of SS has been frequently considered. No one is sure what agents could start this process in Sjögren’s syndrome. Indirect evidence suggest that several viruses may play a role. Research interest has concentrated on the Epstein-Barr virus which has been found in salivary gland biopsies from patient with SS (9). In most cases EBV infects salivary glands leading to a flu-like illness, sometimes with mildly swollen parotid glands. Reactivation of latent EBV occurs more frequently in patients with immunological imbalances or/during other viral infection which temporarily dampen the immune system (6).

Hepatitis C virus may be associated with various autoimmune diseases. Generally mild sialadenitis and a lymphocytic infiltrate in the gland was found in HCV-related liver disease, however, the pathological features were different from SS (41). Interestingly, oral lichen planus, which is now also considered likely to be an autoimmune disease, is often associated with the presence of hepatitis C (20). Some authors even propose that HCV infection should be considered an exclusion criterion for the classification of pSS because the virus may be implicated in the development of SS (42).

Other viruses belonging to the group of retroviruses such as HIV and human T lymphotropic virus type I (HTLV-I) have been suggested as one possible trigger in Sjögren’s syndrome patients (34). Interestingly, 3 month lasting antiretroviral therapy by using reverse transcriptase inhibitors improved both ocular and oral symptoms and also extraglandular manifestation (fatigue, tender points and arthralgias) in all patients with SS participating the study (47). However, the evidence still remains indirect and viral etiology has never been absolutely confirmed.

Nevertheless, the purpose of this review is to summarize recent findings with the focus on the Toll-like receptors, B cells and their possible involvement in pathogenesis of autoimmune disease such as Sjögren’s syndrome.**

### Toll-like receptors

Toll-like receptors (TLR) function as sentinels, alerting the innate immune system in the presence of threatening microbial invasion. They are pattern recognition receptors (PRR), recognizing conserved pathogen-associated molecular patterns (PAMPs), which are expressed by organisms ranging from bacteria, protozoa, fungi to viruses. Till now, 10 different TLRs have been detected in humans and 11 have been identified in mice (48). TLR expression is not confined exclusively to macrophages and DC. They can also be expressed by B cells, NK cells, T cells, mast cells, neutrophils, fibroblasts, epithelial cells, keratinocytes, endothelial cells or cardiac myocytes, respectively (35). TLR3, 7, 8 and 9 are located intracellularly in endosomal compartments, while the rest of TLRs are expressed on the cell surface (31).

TLR2 is essential in the recognition of a wide range of microbial molecules representing groups of species such as Gram-positive and Gram-negative bacteria, as well as mycoplasma and yeast. TLR 2 binds a variety of microbial structures, e.g. peptidoglycan and lipoteichoic acid from Gram-positive bacteria, lipoproteins, lipoarabinomannan from mycobacteria etc. TLR2 can form heterodimers with structurally related TLR1 and TLR6, which facilitates microbial recognition. TLR 1 and 6 participate in discrimination of slight differences between triacyl and diacyl lipopeptides and thus cooperate with TLR 2 (49, 50, 51).

TLR 3 recognizes double-stranded RNA (dsRNA) as well as a synthetic analog of dsRNA, polyinosine-polycytidylic acid (poly(I:C)). dsRNA is a molecular pattern associated with viral infection and activates the synthesis of type I interferons via a MyD88-independent pathway as discussed below (1).

TLR4 is the receptor for Gram-negative lipopolysaccharide (LPS) and is activated by a very small amount of LPS. Furthermore it was found that TLR 4 is implicated in the recognition of endogenous ligands such as heat shock proteins (HSP 60 and HSP 70) (10).

TLR5 is involved in the recognition of flagellin, the major component of the bacterial flagellar filament, from both Gram+ and Gram- bacteria. TLR 5 plays an important role in the recognition of the microbes at the mucosal surface as it is expressed on the intestinal endothelial cells (30).

TLR7 and TLR8 are phylogenetically and structurally related and are involved in the response to viral infection. Human TLR7 recognizes GU-rich short single-stranded RNA (ssRNA) as well as the imidazoquinoline compounds imiquimod (R837) and resiquimod (R848). Human TLR8 is also involved in the recognition of R848 but surprisingly not R837 (14). TLR7 and TLR8 agonists target various cells and differ in cytokine induction profile. TLR7 agonists activate plasmacytoid DCs (pDCs) and B cells leading to the production of IFN-α. TLR8-specific agonists activate myeloid DCs, monocytes and predominantly induce produc-
tion of proinflammatory cytokines and chemokines, such as TNF-α, IL-12 and MIP-1α (11). Interestingly, it has been shown that TLR7/8 are implicated in recognizing ssRNA from diverse viruses such as immunodeficiency and influenza viruses (13, 17).

TLR9 recognizes specific unmethylated CpG-oligodeoxynucleotides (ODN) sequences that distinguish microbial DNA from mammalian DNA. Bacterial DNA contains unmethylated CpG motifs, while in mammalian DNA, the cytosine residues of CpG motifs are highly methylated. In humans, TLR 9 is expressed in B cells and plasmacytoid dendritic cell (PDC), in mice also in the myeloid compartment. Three types of stimulatory ODNs have been described: CpG-A, CpG-B, and CpG-C. CpG-A ODN, also known as 'D'-type, effectively activates NK cells and induces secretion of IFNα in PDC. CpG-B ODN, also known as 'K'-type was identified first and is involved in the activation of B cells. CpG-B ODN are weak inducers of IFNα, however, are very strong Th1 adjuvants and potential B cell response stimulators. CpG-C ODN combines the immune effects of CpG-A and B ODN and trigger B-cell activation similarly to CpG-B together with IFNα secretion as CpG-A ODN. In addition, NK cells stimulated with CpG-C ODN showed higher cytolytic activity against tumor cell lines. Combination of both immune effects might have broader therapeutic consequences in various tumor types and autoimmune diseases (2, 25).

Binding of TLR's ligands triggers signalling via the adaptor protein MyD88, with activation of transcription factors NF-κB and AP-1, resulting in secretion of inflammatory cytokines such as TNFα and IL 12p40 and upregulation of co-stimulatory molecules (49). Different signalling pathways either dependent or independent on MyD88 were described. In the MyD88-dependent pathway, MyD88 adaptor protein plays a pivotal role. It was shown that MyD88 is used by all TLRs, supported by MyD88-deficient mice model, where no inflammatory cytokines were produced in response to TLR ligands (52, 21). Although members of the TLR signal through MyD88, the signalling pathways induced by individual receptors differ. Another adaptor molecule, which is structurally related to MyD88, was identified and termed as TRAM/TICAM-2. Studies in mice showed that TRAM is implicated in TLR 4-mediated activation of IFN-3 and induction of type I INF expression (38). In conclusion, TRAM is important for TRIF dependent/MyD88-independent pathway via TLR 4.

TLRs have a key function in triggering adaptive immune responses to microbes and are probably implicated in inflammatory and autoimmune disorders. Involvement of the TLR-9-MyD88-dependent pathway in the induction of SLE and RA has been described by several research groups (53, 5). Thus, innate immunity plays a fundamental role in the initiation of the immune reactions.

**B cell Toll like receptors**

An advance in understanding of autoimmune processes has led to the clue, that B cells have additional functions, besides autoantibody secretion. It looks that B cells can present antigen, secrete cytokines and regulate T cells activities independently of antibody production. Thus, application of B cell depletion therapy using rituximab, a chimeric anti-CD20 antibody, has improved condition of SLE and RA. Although it would seem that the effect arises from the elimination of autoantibody-secreting B cells, many patients in clinical remission had no decline in serum levels of relevant autoantibodies (45, 39).

Various B cells subsets secrete both proinflammatory (TNFα, LTα, IL-12, IL-6) and suppressive (IL-10) cytokines. The cytokine profile of B cells can also be disturbed by disease, leading to an imbalance of proinflammatory and anti-inflammatory cytokines (29). In patients with multiple sclerosis (MS), the cytokine network was dysregulated. It was evidenced that in MS patients the expression of anti-inflammatory cytokine IL-10 was decreased while the treatment with rituximab reversed the ratio in favor to anti-inflammatory cytokines (8).

Experimental data support the idea that autologous nucleic acids, perhaps from necrotic or apoptotic cells, in some circumstances may be stimulatory ligands for TLR7 and TLR9. B cells with BCR specific for antigens complexed with nucleic acids (or specific for the nucleic acids themselves) may internalize the complexes by receptor-assisted endocytosis, making the nucleic acids available for binding to endosome-associated TLR7 or TLR9. Thus, these complexes may be both autoantigens and autoadjuvants (36, 35). Indeed, evidence has been presented that engagement of B cell TLR can trigger a humoral immune response. The crucial insight into this problem, demonstrating involvement of BCR and TLR signals in autoantibody production, came from Ann Marshak Rothstein's group. In a model with B cells from AM14 transgenic mice,
complexes of chromatin and IgG2a induced production of rheumatoid factor by dual engagement of BCR and TLR9 (28). Moreover in the absence of MyD88, TLR 7 and TLR 9, proliferation of AM14 B cells was abolished (54). Later, a similar effect was reported for TLR7. In this case, the response was more vigorous when IFNα was added. IFNα upregulates TLR7, and is often increased during infections (27).

Some studies have started to deal in more detail with this issue. Study by Han et al. compare the effect of TLR7 and 9 on human B cells function by determining gene expression and protein production of several chemokine, cytokine and B cell activation markers. TLR7, 7/8, 9 agonists directly induced the expression of co-stimulatory molecules including CD80, CD86, CD40, CD58 in a similar fashion. Additionally two receptors were modulated by TLR7. 7/8, 9 agonists. CD23, an important molecule for B cell activation was up regulated, while CD32, resulting in inhibition of B cells was down regulated. The study has demonstrated that B cells were activated by agonists of TLR7 and TLR7/8 in a similar way to the TLR9 agonist (46).

Interestingly, Bourke et al. systematically investigated 10 TLRs mRNA expression profile in normal B cells obtained from human tonsils and malignant B cells including pre-B cell, Epstein-Barr virus (EBV)-transformed cell lines, Burkitt lymphoma and other neoplastic cell lines. The results have demonstrated that TLR9 and TLR10 were predominantly expressed on human B lymphocytes, similarly to high expression of TLR9 and TLR10 in EBV-transformed B-cell lines, Burkitt lymphoma, follicular lymphoma, and multiple myeloma cell lines. However, pre-B cell lines were negative for TLR9 and TLR10, supporting the theory that expression of TLR9 and TLR10 correlates with B-cell differentiation and maturation. In result, stimulation through TLR may create a link between chronic infection and pathogenesis of lymphomas (4).

Shlomchik (46) proposed a model of autoimmunity feedback cycle, where the break of tolerance is initiated by recognition of auto-Ags that carry an endogenous TLR ligand such as DNA. Ligation of BCR and TLR leads to presentation of auto-Ag to autoreactive T cells and activation of T cells with expression of CD40 and IL-21 along with other costimulatory molecules and cytokines. Activated T cell may then broaden previous autoreactive B cell reaction and help them to enhance clonal expansion, isotype switch, somatic hypermutation, affinity maturation and differentiation into antibody-secreting cells and activation of many other autoreactive B cells.

It seems that engagement of BCR and TLR might be the initial point in breaking tolerance, and the presence of T cells is required subsequently to generate a fully-developed autoimmune response. However, T cell tolerance must be also breached. The activation of autoreactive T cells can be initiated though APCs, such as DCs and macrophages, which are able to activate naive T cells. On the other hand, autoreactive B cells that have previously been activated in T cell-independent/BCR and TLR-dependent way, can activate autoreactive T cells resulting in boosting and terminal development of autoimmune reaction.

**Naive and memory B cells**

Several animal studies have indicated that there is a synergism between TLR and B cells (28). The crosslinking of TLR and BCR in the presence of antigens containing polyclonal activator, such as LPS or unmethylated DNA, might be a critical step in pathogenesis of autoimmune diseases such as systemic lupus erythematosus (24). Mouse naive B cells, compared to human naive B cells, express high levels of TLR9 and can be activated by TLR ligands (LPS or CpG) without additional help from T cells or BCR stimulation. In humans, TLRs are expressed at very low levels by naive B cells, until they are stimulated through BCR. In contrast to human naive B cells, human memory B cells constitutively express high levels of TLR2, -6, -7, -9 and -10 (26, 36). Triggering via TLR 7 and 9 agonists leads to proliferation and differentiation of human memory B cells into Ig-secreting cells, cytokine production and up-regulation of activation markers for antigen presentation to (autoreactive) T cells (36). In addition, human memory B cells can be activated with the help of T cells, independently of an antigen (26). Bernasconi et al. proposed that the upregulation of TLR9 expression following BCR triggering cooperates in the activation of naive B cells. In the first step antigen binds to BCR and triggers TLR9 expression followed by internalization of antigen with its TLR9 ligand in the endosomal compartment and interaction with newly synthesized TLR9. CpG DNA has been considered to be a potent stimulant of B cells, enhancing final differentiation. After stimulation of naive and memory B cell subsets with CpG, anti-Ig and various cytokines, several observations were made. The response of naive and memory B cells did differ. Naive B cells proliferated only when stimulated by CpG, anti-Ig, IL-2 and IL-10, while there was no significant difference in the response of memory B cells either in the presence or in the absence of anti-Ig. It was also shown that naive and memory B cells proliferation can be induce in the T independent fashion (3).

This makes the human immune system highly specific, because coupling of TLR 9 and BCR in the absence of T cells enables the naive B cells to be the only ones activated by microbial stimuli. In contrast, constitutive expression of TLR in memory B cells ensures permanent support in antibody production of all memory specificities thereby sustaining serological memory. Thus, a novel role for TLRs in the immune response was suggested (36, 46, 26, 3).

More recently, Ruprecht and Lanzavecchia (44) proposed a model of activation of human naive B cells. According to their findings it seems that three signals are required for optimal activation: Antigen binding to BCR (signal one), specific binding of T cell receptor (signal two) and engagement of a TLR (signal three). It was shown that
signal one and two are sufficient for starting initial proliferation of naive B cells, however, they fail to keep B cell expansion and proliferation. In one of the experiments, naive B cells were stimulated with anti-IgG, T cell help (CD4+ T cells in presence of bacterial superantigen TSST) and CpG. After 24 hours all stimuli were removed and then repeatedly added alone or in combination. When added CpG and T cell help alone, the response was only partly re-established. The effective response was reached when stimuli were given together. In conclusion, TLR stimulation, as a third signal, is a critical point in activation of human naive B cells.

A few clinical studies with the focus on Toll like receptors of SS patients exist. Zheng et al. investigated the expression of TLR 7, 8, 9 in peripheral mononuclear blood cells (PMBCs) and the existence of TLR 7 and 9 in minor parotid glands in patients with primary Sjögren’s syndrome (pSS). They found that expression of TLR 7 and 9 was upregulated in pSS, while TLR 8 was not. Moreover, TLR 7 and 9 positive cells were detected in the epithelial islands, lymphocytes and ductal epithelial cells of the parotid glands in pSS, and were more frequent than in controls (57). Interestingly, another study by Zheng et al. focused on interferon-regulatory factors (IRF) of the Toll-like receptor family. Expression of IRF 1, 3, and 7 in PMBCs was investigated, and upregulated levels of IRF 1 in patients with pSS compared to controls was demonstrated (58). Kawakami et al. investigated expression of TLRs in labial salivary glands of patients with SS, where TLR 2, 3, 4 and MyD88 were more strongly expressed than in controls and involvement of mitogen-activated protein kinase pathway was suggested (23). Interesting findings came from Manoussakis et al., who focused on TLR 3 expression and possible link to the apoptotic death. In this study, polyI:C-treated salivary gland epithelial cells were found to suffer detachment-induced apoptosis, so-called anoikis. Pro-apoptotic molecules such as Bmf, BimEL and Bax were upregulated, while pro-survival Bcl-2 was downregulated. Finally, it was found that salivary gland epithelial cells derived from pSS patients were particularly susceptible to TLR-3-induced anoikis than in control group. These activation processes may operate in the epithelial cells of SS patients (32).

### Concluding remarks

Experimental evidence and clinical results demonstrate a key role for B cell and Toll like receptors, mainly TLR 7, 9 and 3, in the development of systemic autoimmune disease.

### Tab. 1: Summary of the characteristics of TLRs and their expression on human naive and memory B cells.

<table>
<thead>
<tr>
<th>TLR</th>
<th>ligands</th>
<th>cellular location</th>
<th>adaptor protein</th>
<th>expression of TLR on human memory B cells</th>
<th>expression of TLR on human naive B cells</th>
</tr>
</thead>
<tbody>
<tr>
<td>TLR-1, 2, 6</td>
<td>lipoproteins, peptidoglycans lipoteichoic acids, zymosan, LPS</td>
<td>cell surface</td>
<td>MyD88, TIRAP</td>
<td>constitutively high level</td>
<td>not detected</td>
</tr>
<tr>
<td>TLR-4</td>
<td>LPS, HSP60/70</td>
<td>cell surface</td>
<td>MyD88, TIRAP, TRAM, TRIF</td>
<td>not detected</td>
<td>not detected</td>
</tr>
<tr>
<td>TLR-5</td>
<td>flagellin</td>
<td>cell surface</td>
<td>MyD88</td>
<td>not detected</td>
<td>not detected</td>
</tr>
<tr>
<td>TLR-3</td>
<td>dsRNA</td>
<td>endosomal compartment</td>
<td>TRIF</td>
<td>not detected</td>
<td>not detected</td>
</tr>
<tr>
<td>TLR-7/8</td>
<td>ssRNA</td>
<td>endosomal compartment</td>
<td>MyD88</td>
<td>constitutively high level, upregulated through BCR stimulation</td>
<td>upregulated through BCR stimulation</td>
</tr>
<tr>
<td>TLR-9</td>
<td>unmethylated CpG DNA</td>
<td>endosomal compartment</td>
<td>MyD88</td>
<td>constitutively high level, upregulated through BCR stimulation</td>
<td>upregulated through BCR stimulation</td>
</tr>
</tbody>
</table>

Legend: MyD88: Myeloid differentiation primary response protein 88; TIRAP: Toll/IL-1 receptor-domain-containing adaptor protein; TRIF: Toll/IL-1 receptor-domain-containing adaptor protein inducing INF-β; TRAM: TRIF-related adaptor molecule; dsRNA: Double-stranded RNA; ssRNA: Single-stranded RNA; HSP: Heat shock protein
seases such as SS, SLE, RA. An overview of hypothetical involvement B cell TLRs is shown in Fig. 1. However, in order to fully understand the exact mechanism leading to SS, further studies focusing on the subject are required. This may provide detailed insight into the pathogenesis of autoimmune disease and could be helpful for future development of a novel therapy.

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Fig. 1: Three signals model for the induction of the autoimmunity.

Legend: Combined activation through the TLR 7 and 9, the induction of autoantigen to autoreactive BCR and help provided by TH2 T cells might be necessary for triggering autoimmune reaction. Activation of autoreactive B cells leads to efficient proliferation of B cell, their terminal differentiation into plasma cells and production of large amount of autoantibodies. In addition, this results in somatic hypermutation, ultimating into the hyperaffinity antibodies formation, isotype switching and in the last to the immunological memory formation.

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